AMENDMENTS TO THE SPECIFICATION

Please replace the paragraph beginning on page 26, line 1 with the following rewritten paragraph:

Fusion polypeptides between 312C2s and other homologous or heterologous proteins are also provided. Many cytokine receptors or other surface proteins are multimeric, e.g., homodimeric entities, and a repeat construct may have various advantages, including lessened susceptibility to proteolytic cleavage. Typical examples are fusions of a reporter polypeptide, e.g., luciferase, with a segment or domain of a protein, e.g., a receptor-binding segment, so that the presence or location of the fused ligand may be easily determined. See, e.g., Dull, et al., U.S. Patent No. 4,859,609. Other gene fusion partners include bacterial ß-galactosidase, trpE, Protein A, ß-lactamase, alpha amylase, alcohol dehydrogenase, yeast alpha mating factor, and detection or purification tags such as a FLAG sequence of His6 sequence (SEQ ID NO:9). See e.g., Godowski, et al. (1988) Science 241:812-816.

Please replace the paragraph beginning on page 26, line 31 with the following rewritten paragraph:

This invention also contemplates the use of derivatives of 312C2s other than variations in amino acid sequence or glycosylation. Such derivatives may involve covalent or aggregative association with chemical moieties. Covalent or aggregative derivatives will be useful as immunogens, as reagents in immunoassays, or in purification methods such as for affinity purification of binding partners, e.g., other antigens. A 312C2 can be immobilized by covalent bonding to a solid support such as cyanogen bromide-activated ~~SEPHAROSE~~ SEPHAROSE®, by methods which are well known in the art, or adsorbed onto polyolefin surfaces, with or without glutaraldehyde cross-linking, for use in the assay or purification of anti-312C2 antibodies or an alternative binding composition. Western blot techniques are also common. The 312C2s can also be labeled with a detectable group, e.g., for use in diagnostic assays. Purification of 312C2 may be effected by an immobilized antibody or complementary binding partner.

Please replace the paragraph beginning on page 56, line 21 with the following rewritten paragraph:

The PCR-based subtraction system developed by Wang and Brown (1991) Proc. Natl. Acad. Sci. USA 88:11505-11509, was modified to apply to plasmid cDNA libraries. A cDNA library specific for activated αβDN thymocytes was generated using 100 μg of the unstimulated αβDN cDNA library DNA digested with XbaI, NotI, and ScaI as driver DNA and 5 μg of the stimulated αβDN cDNA library DNA as tracer DNA. Following restriction digestion, the driver DNA was treated with DNA polymerase Klenow fragment to fill-in the restriction sites. After ethanol precipitation, the DNA was dissolved in 100 μl of water, heat-denatured and mixed with 100 μl (100 μg) of ~~Photoprobe~~ PHOTOPROBE® biotin (Vector Laboratories, Burlingame, CA). The driver DNA was then irradiated with a 270-W sunlamp on ice for 20 min. 50 μl more ~~Photoprobe~~ PHOTOPROBE® biotin was added and the biotinylation reaction was repeated. After butanol extraction, the photobiotinylated DNA (driver-U) was ethanol-precipitated and dissolved in 30 μl of 10 mM Tris-HCl and 1 mM EDTA, pH 8 (TE). As tracer DNA, 5 μg of stimulated αβDN cDNA was digested with XbaI and NotI; ethanol precipitated; and dissolved in 4 μl of TE (tracer-S). Tracer-S was mixed with 15 μl of driver-U, 1 μl (10 μg) of E. coli tRNA (Sigma, St. Louis, MO), and 20 μl of 2 x hybridization buffer (1.5 M NaCl, 10 mM EDTA, 50 mM HEPES, pH 7.5, 0.2% SDS), overlaid with mineral oil, and heat-denatured. The sample tube was immediately transferred into a 68° C water bath and incubated for 20 h. The reaction mixture was then subjected to streptavidin treatment followed by phenol/chloroform extraction. Subtracted DNA was precipitated, dissolved in 12 μl of TE, mixed with 8 μl of driver-U and 20 μl of 2 x hybridization buffer, and then incubated at 68° C for 2 h. After streptavidin treatment, the remaining DNA was ligated with 250 ng of a purified XbaI / NotI fragment of pJFE-14 and then transformed into electro-competent E. coli cells to generate the activation specific αβDN subtracted library (S1). 100 independent clones were randomly picked and screened by hybridization using a cocktail of known cytokine cDNAs. Plasmid DNA's were prepared from clones that did not hybridize to the cytokine probes. These clones were grouped by insert size and further characterized by DNA sequencing. Clones corresponding to the 312C2 were isolated.